

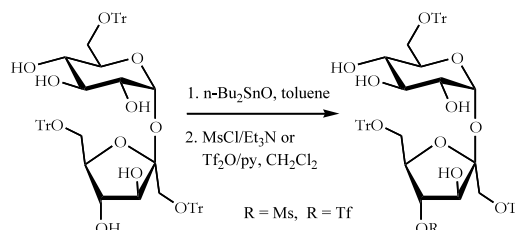
Regioselective sulfonylation of 6,1',6'-tri-O-tritylsucrose through dibutylstannylation: synthesis of 4'-O-sulfonyl derivatives of sucrose

Carbohydr. Res. **2002**, *337*, 2377

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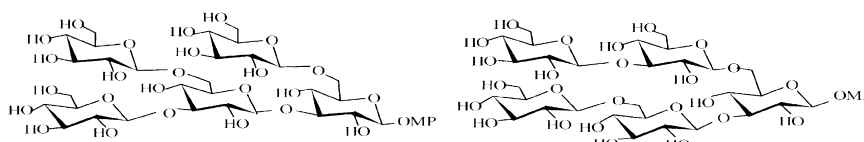


Synthesis of two isomeric pentasaccharides, the possible repeating unit of the β -glucan from the micro fungus *Epicoccum nigrum* Ehrenb. ex Schlecht

Carbohydr. Res. **2002**, *337*, 2383

Ying Zeng, Wenhui Zhang, Jun Ning, Fanzuo Kong

Research Center for Eco-Environmental Sciences, Academia Sinica, PO Box 2871, Beijing 100085, China

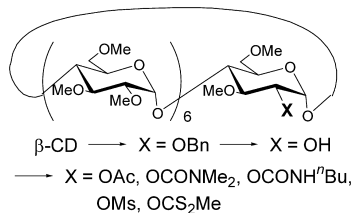


Facile preparation of mono-2-O-modified eicosa-O-methylcyclomaltoheptaoses (β -cyclodextrins)

Carbohydr. Res. **2002**, *337*, 2393

Masato Suzuki, Yutaka Nozoe

Department of Organic and Polymeric Materials, Graduate School of Science and Engineering, and International Research Center of Macromolecular Science, Tokyo Institute of Technology, 2-12-1 O-okayama, Meguro-ku, Tokyo 152-8552, Japan



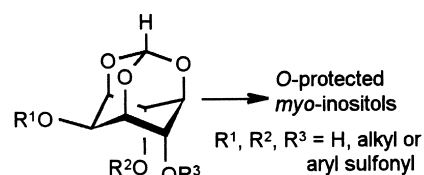
Sulfonate protecting groups. Regioselective sulfonylation of *myo*-inositol orthoesters—improved synthesis of precursors of D- and L-*myo*-inositol 1,3,4,5-tetrakisphosphate, *myo*-inositol 1,3,4,5,6-pentakisphosphate and related derivatives

Carbohydr. Res. **2002**, *337*, 2399

Kana M. Sureshan,^a Mysore S. Shashidhar,^a Thoniyot Praveen,^a Rajesh G. Gonnade,^b Mohan M. Bhadbhade^b

^aDivision of Organic Synthesis, National Chemical Laboratory, Pune 411 008, India

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Novel amphiphilic fluoroalkylated derivatives of xylitol, D-glucose and D-galactose for medical applications: hemocompatibility and co-emulsifying properties

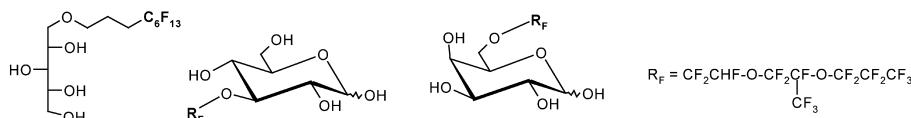
Carbohydr. Res. **2002**, *337*, 2411

Oldřich Paleta,^a Ivona Dlouhá,^a Robert Kaplánek,^a Karel Kefurt,^b Milan Kodíček^c

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^bDepartment of Natural Compounds Chemistry, Prague Institute of Chemical Technology, Technická 5, 16628 Prague 6, Czech Republic

^cDepartment of Biochemistry and Microbiology, Prague Institute of Chemical Technology, Technická 5, 16628 Prague 6, Czech Republic



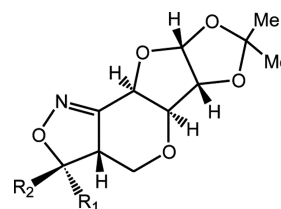
Synthesis, high-resolution NMR spectroscopic analysis, and single-crystal X-ray diffraction of isoxazoline tetracycles

Carbohydr. Res. **2002**, *337*, 2419

Mirta L. Fascio,^a Angel Alvarez-Larena,^b Norma B. D'Accorso^a

^aCentro de Investigaciones de Hidratos de Carbono (CIHIDECAR), Departamento de Química Orgánica, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Ciudad Universitaria, Pabellón II, 3° Piso, C.P. 1428, Buenos Aires, Argentina

^bUnitat de Cristal·lografia, Universitat Autònoma de Barcelona, 08193 Bellaterra, Barcelona, Spain



4 R₁ = H; R₂ = CH₃

5 R₁ = H; R₂ = Ph

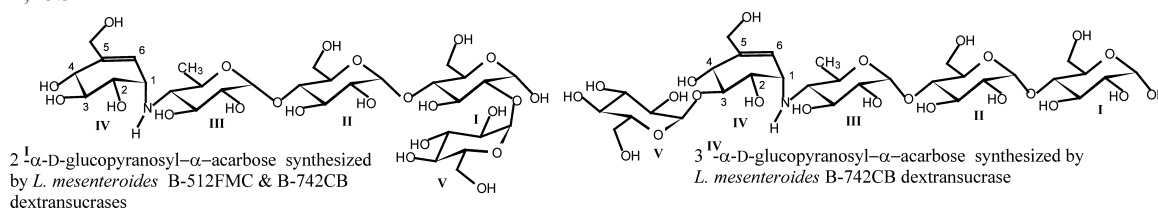
6 R₁ = CH₃; R₂ = CH₃

Synthesis of acarbose analogues by transglycosylation reactions of *Leuconostoc mesenteroides* B-512FMC and B-742CB dextranases

Carbohydr. Res. **2002**, *337*, 2427

Seung-Heon Yoon, John F. Robyt

Laboratory of Carbohydrate Chemistry and Enzymology, 4252 Molecular Biology BLDG, Iowa State University, Ames, IA 50011, USA



The binding of synthetic analogs of the upstream, terminal residue of the O-polysaccharides (O-PS) of *Vibrio cholerae* O:1 serotypes Ogawa and Inaba to two murine monoclonal antibodies (MAbs) specific for the Ogawa lipopolysaccharide (LPS)

Carbohydr. Res. **2002**, *337*, 2437

Ximan Liao,^a Emanuel Poirot,^a Alex H. C. Chang,^a Xiaodong Zhang,^a Jian Zhang,^a Farida Nato,^b Jean-Michel Fournier,^b Pavol Kováč,^a Cornelis P. J. Glaudemans^a

^aLaboratory of Medicinal Chemistry, NIDDK, National Institutes of Health, Bethesda, MD 20892-0815, USA

^bUnité du Choléra et des Vibrions, Centre National des Référence des Vibrions et du Choléra, and Laboratoire d'Ingénierie des Anticorps, Institut Pasteur, F-75724 Paris Cedex 15, France

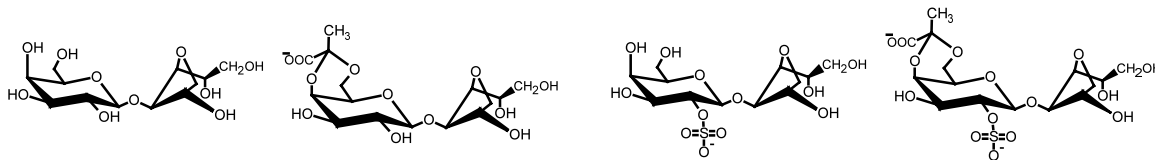
The refined hydrogen-bonding pattern involved in the title antigen–antibody system is reported.

Sulfated and pyruvylated disaccharide alditols obtained from a red seaweed galactan: ESIMS and NMR approaches

Carbohydr. Res. **2002**, *337*, 2443

Alan G. Gonçalves, Diogo R. B. Ducatti, M. Eugênia R. Duarte, Miguel D. Nosedá

Departamento de Bioquímica e Biologia Molecular, Universidade Federal do Paraná, PO Box 19046, Curitiba, Paraná, Brazil



Preparation and characterisation of chitosans with oligosaccharide branches

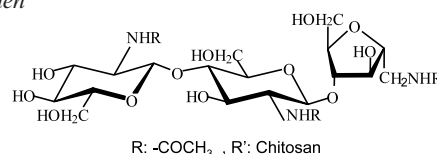
Carbohydr. Res. **2002**, *337*, 2455

Kristoffer Tømmeraas,^a Magnus Köping-Höggård,^b Kjell M. Vårum,^a Bjørn E. Christensen,^a Per Artursson,^b Olav Smidsrød^a

^aNorwegian Biopolymer Laboratory (NOBIPOL), Department of Biotechnology, Norwegian University of Science and Technology (NTNU), N-7491 Trondheim, Norway

^bDepartment of Pharmacy, Uppsala University, Box 580, SE-751 23 Uppsala, Sweden

2-Acetamido-2-deoxy-D-glucopyranosyl-β-(1→4)-2-acetamido-2-deoxy-D-glucopyranosyl-β-(1→4)-2,5-anhydro-D-mannofuranose (A–A–M) was reductively *N*-alkylated onto a fully de-*N*-acetylated chitosan ($F_A < 0.001$, $DP_n = 25$) to obtain branched chitosans with degree of substitution (DS) of 0.070, 0.23 and 0.40 which were characterised.



Structure of the O-specific polysaccharide of *Proteus vulgaris* O15 containing a novel regioisomer of *N*-acetylmuramic acid, 2-acetamido-4-*O*-[(*R*)-1-carboxyethyl]-2-deoxy-D-glucose

Carbohydr. Res. **2002**, *337*, 2463

Andrei V. Perepelov,^a Agnieszka Torzewska,^b Alexander S. Shashkov,^a Andrzej Ziolkowski,^c Sof'ya N. Senchenkova,^a Antoni Rozalski,^b Yuriy A. Knirel^a

^aN. D. Zelinsky Institute of Organic Chemistry, Russian Academy of Sciences, 119991 Moscow, Russian Federation

^bDepartment of Immunobiology of Bacteria, Institute of Microbiology and Immunology, University of Lodz, 90-237 Lodz, Poland

^cCentre of Microbiology and Virology, Polish Academy of Sciences, Lodowa 106, 93-232 Lodz, Poland

The polysaccharide was shown to have the following repeating unit:

→3)-α-D-GlcpNAc4(*R*-Lac)6Ac-(1→2)-β-D-GlcpA-(1→3)-α-L-6dTalp2Ac-(1→3)-β-D-GlcpNAc-(1→ where L-6dTalp and D-GlcpNAc4(*R*-Lac) are 6-deoxy-L-talose and 2-acetamido-4-*O*-[(*R*)-1-carboxyethyl]-2-deoxy-D-glucose, respectively.

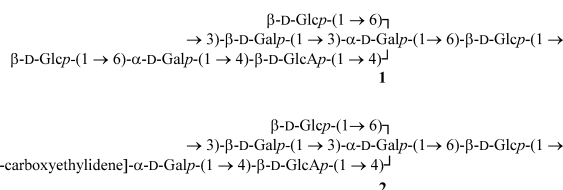
Extracellular polysaccharides of *Erwinia futululu*, a bacterium associated with a fungal canker disease of *Eucalyptus* spp.

Carbohydr. Res. **2002**, *337*, 2469

Byung Yun Yang, Qiong Ding, Rex Montgomery

Department of Biochemistry, College of Medicine, University of Iowa, Iowa City, IA 52242, USA

Extracellular polysaccharides (EPSs) produced by an *Erwinia* spp. associated with a fungal canker disease of *Eucalyptus* were fractionated into two polysaccharides, one (1) that was identified with that produced by *Erwinia stewartii*. The other (2) has a similar structure, but with one terminal Glc residue replaced by pyruvic acid to give 4,6-*O*-(1-carboxyethylidene)-Galp.

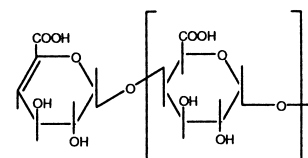
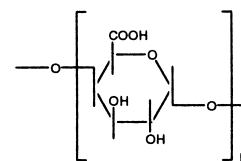


High-performance liquid chromatographic separation and on-line mass spectrometric detection of saturated and unsaturated oligogalacturonic acids

Thomas Stoll, Andreas Schieber, Reinhold Carle

Institute of Food Technology, Section Plant Foodstuff Technology, Hohenheim University, Garbenstraße 25, D-70599 Stuttgart, Germany

An analytical system for the simultaneous separation of saturated and unsaturated oligogalacturonic acids (up to dp 7) is presented allowing on-line mass spectrometric detection without additional desalting.



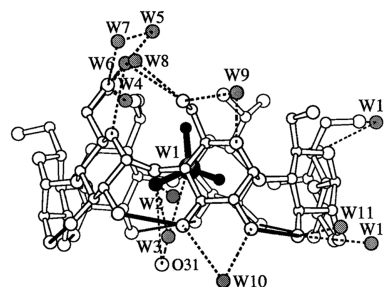
Crystal structure of β -cyclodextrin–dimethylsulfoxide inclusion complex

Thammarat Aree,^a Narongsak Chaichit^b

^a*Department of Chemistry, Faculty of Science, Chulalongkorn University, Phiyathai Road, Pathumwan, Bangkok 10330, Thailand*

^b*Department of Physics, Faculty of Science and Technology, Thammasat University, Rangsit, Pathum Thani 12121, Thailand*

Crystallographic evidence for a β -CD–DMSO inclusion complex is presented. DMSO is entirely embedded in the β -CD cavity and is maintained in position by hydrogen bonding to the water site W-3 and O-31–H group of a symmetry equivalent β -CD.



Preparation and investigation of antibacterial carbohydrate-based surfaces

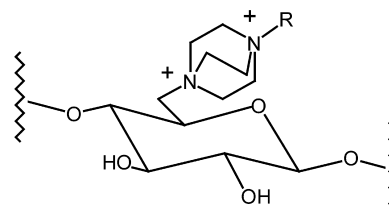
Tanya Abel,^a JaimeLee Iolani Cohen,^a Robert Engel,^c
Maya Filshtinskaya,^a Alice Melkonian,^b Karin Melkonian^b

^a*Department of Chemistry and Physical Sciences, Pace University, 1 Pace Plaza, W-333, New York, NY 10038, USA*

^b*Department of Biology, Long Island University, C.W. Post Campus, Northern Boulevard, Greenville, NY 11548, USA*

^c*Department of Chemistry and Biochemistry, Queens College of CUNY and the Graduate Center of CUNY, 65-30 Kissena Boulevard, Flushing, NY 11367, USA*

Surfaces bearing carbohydrate units have been modified in a two-step process to incorporate functionalities (lipophilic with polycationic units) that bear antibacterial activity. The effectiveness of these modified surfaces for antibacterial action against a series of seven Gram-positive and Gram-negative bacteria are reported.



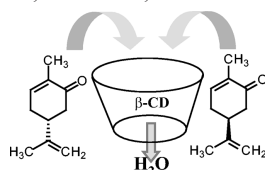
Inclusion of carvone enantiomers in cyclomaltoheptaose (β -cyclodextrin): thermal behaviour and H \rightarrow D and D \rightarrow H exchange

Aida M. Moreira da Silva,^a José M. A. Empis,^b José J. C. Teixeira-Dias^c

^a*Department of Food Science and Technology, ESAC, Bencanta, P-3040 Coimbra, Portugal*

^b*Department of Chemical Engineering, Instituto Superior Técnico, P-1096 Lisboa, Portugal*

^c*Department of Chemistry, University of Aveiro, CICECO, P-3810 Aveiro, Portugal*



Structure of the oligomers obtained by enzymatic hydrolysis of the glucomannan produced by the plant *Amorphophallus konjac*

Paola Cescutti, Cristiana Campa, Franco Delben, Roberto Rizzo

Dipartimento di Biochimica, Biofisica e Chimica delle Macromolecole, Università di Trieste, via L. Giorgieri 1, I-34127 Trieste, Italy

Dimers and trimers obtained by enzymatic hydrolysis of konjac glucomannan were characterised by means of electrospray mass spectrometry, capillary electrophoresis and NMR. The investigation revealed that the polysaccharidic chain is composed of random sequences of Glc and Man residues.